

# myReads® Sample Drying Guide

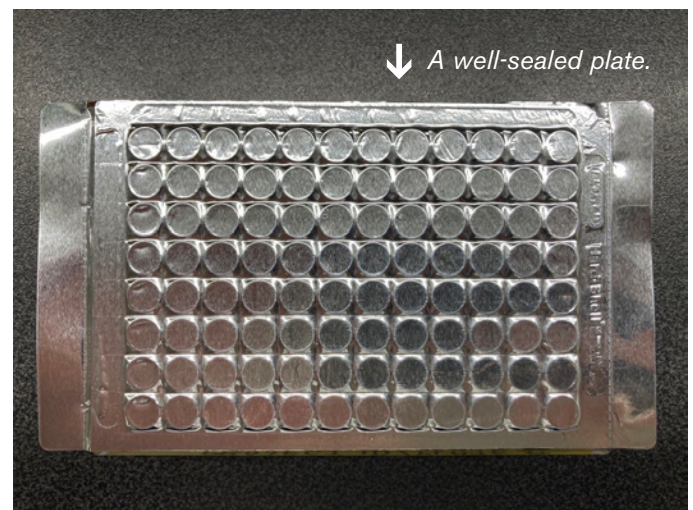
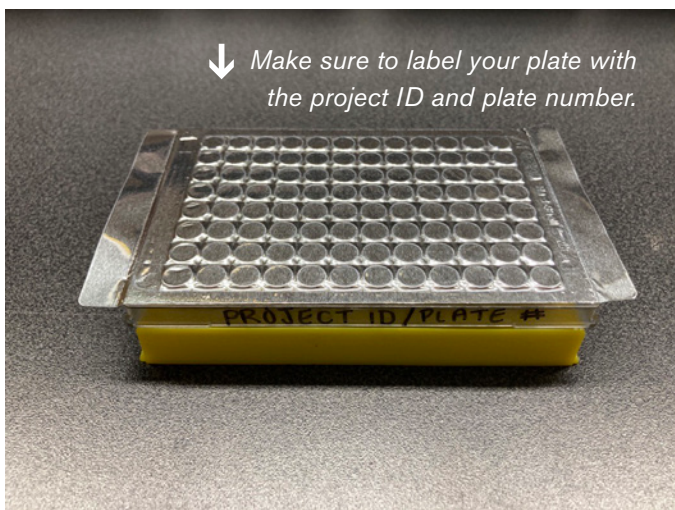
**Please follow the instructions below carefully when preparing your DNA samples for submission.**

Samples which do not meet these requirements will not be processed and may incur additional fees.

**What:** All DNA submissions are required to be fully dried and in plate format.

**Why:** Samples that are in liquid format (even if frozen) are vulnerable to thawing, and potentially splashing, resulting in cross contamination. If samples are in liquid format and a well is crushed during the journey, the samples will leak out and be lost. With dried samples, there is a chance for recovery from a damaged tube/plate. Dried nucleic acids are typically stable at room temperature for several months and can survive an extended delay due to customs holds/shipment errors/etc. The reason we ask for samples to be dried is because we want them to arrive safely!

**How:** Below, we suggest three options for drying . Other methods are acceptable as long as your plate is full- or semi-skirted and your seal is an adhesive foil. (*Pro-tip – make sure you press thoroughly around and between the wells when you seal the plate!*)



Suggested plastics/consumables:

- 96-well clear semi-skirted plate: Bio-Rad, catalog number HSS9601 | [Purchase Here](#)
- Adhesive foil seal: VWR, catalog number 60941-076 | [Purchase Here](#)

## Three Options for Drying

### 1. Vacuum centrifugation (vacuum concentration):

- *Suggested vacuum concentrator protocol:* Time for drying will vary based on starting volume and equipment. For standard/fresh samples: Preheat a vacuum centrifuge to high heat (60C). Set the plate into the vacuum centrifuge without a plate seal and start the protocol, seal the plate when the liquid has evaporated. Check every 15 minutes to monitor progress. For heavily degraded/ancient samples: Set vacuum centrifuge to no heat, or lowest possible heat. Set the plate into the vacuum centrifuge without a plate seal and start the protocol, seal the plate when the liquid has evaporated. Check every 30 minutes to monitor progress.
- *Pros & cons:* Fast, but requires special equipment.

### 2. Passive drying:

- *Suggested passive drying protocol:* Leave the plate open in a gently heated (~37°C) thermal cycler until the liquid has evaporated. Seal plate. (*Pro-tip – make sure no bugs or bits of fluff fall into the wells during drying!*)
- *Pros & cons:* No special equipment needed, but slow.

### 3. Bind to a silica membrane:

- *Suggested silica membrane binding protocol:* Follow the protocol of the kit through the binding, washing, and drying steps, but do not perform the final elution, just seal the plate instead. Send myReads the specifications for volume and type of elution buffer.
- *Pros & cons:* No special equipment required, but silica membrane plates are more expensive than regular plates and purification always results in some loss of gDNA mass.
- *Suggested kits:*
  - Standard: QIAquick 96 PCR Purification Kit, Qiagen, catalog number 28181 | [Purchase Here](#)
  - Highly Degraded: QIAquick Nucleotide Removal Kit, Qiagen, catalog number 28306 | [Purchase Here](#)

**What if I don't comply?** You have two options: 1) we reseal your shipping box and ship it right back to you, at your cost; or 2) you pay a \$500/plate reformatting fee, which must be paid before we touch your samples.

**Questions?** Email us at [service@arbor.daicel.com](mailto:service@arbor.daicel.com).

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